

Supplementary Information for “multiSLIDE is a web server for exploring connected elements of biological pathways in multi-omics data”

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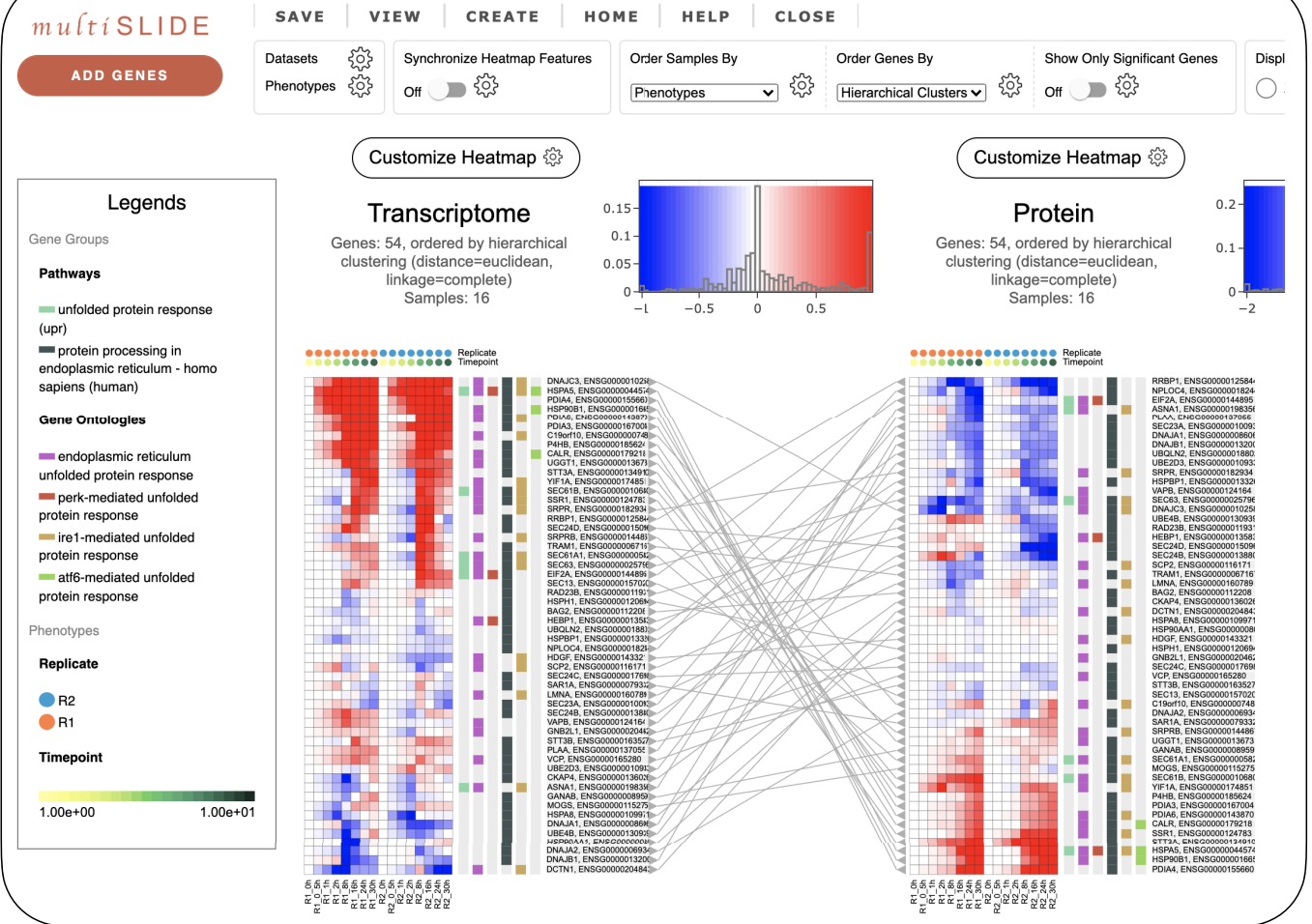
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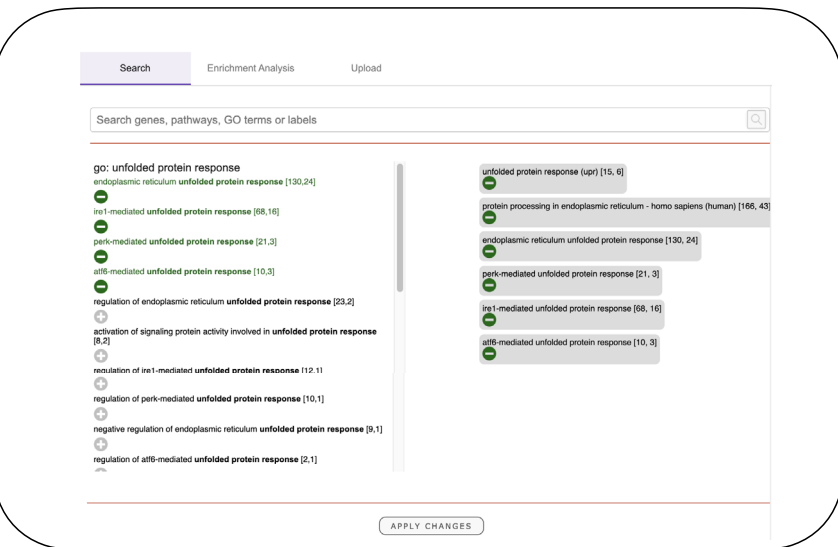
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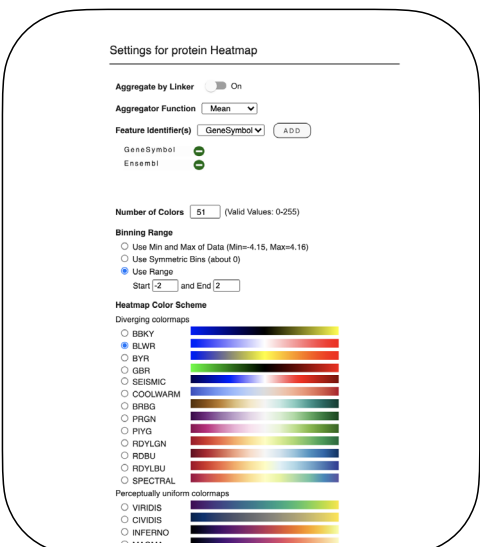
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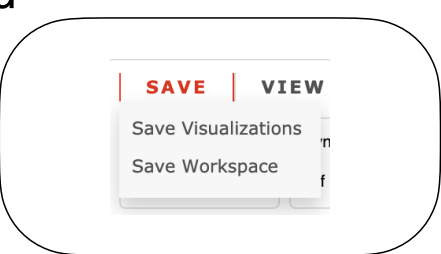
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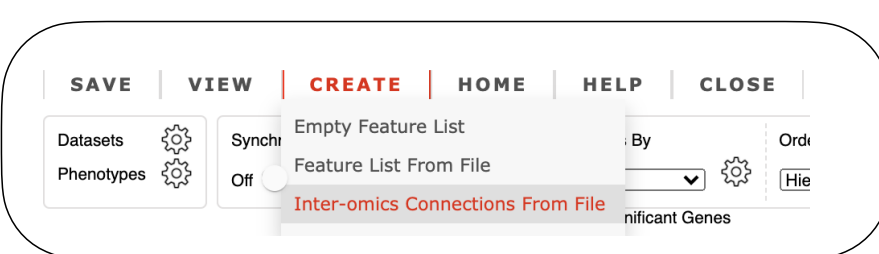
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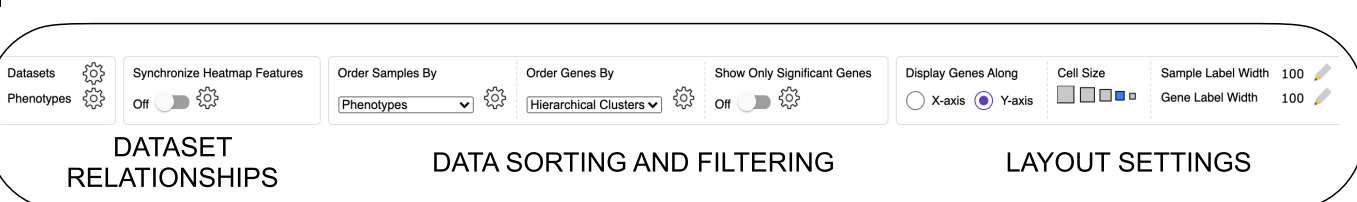
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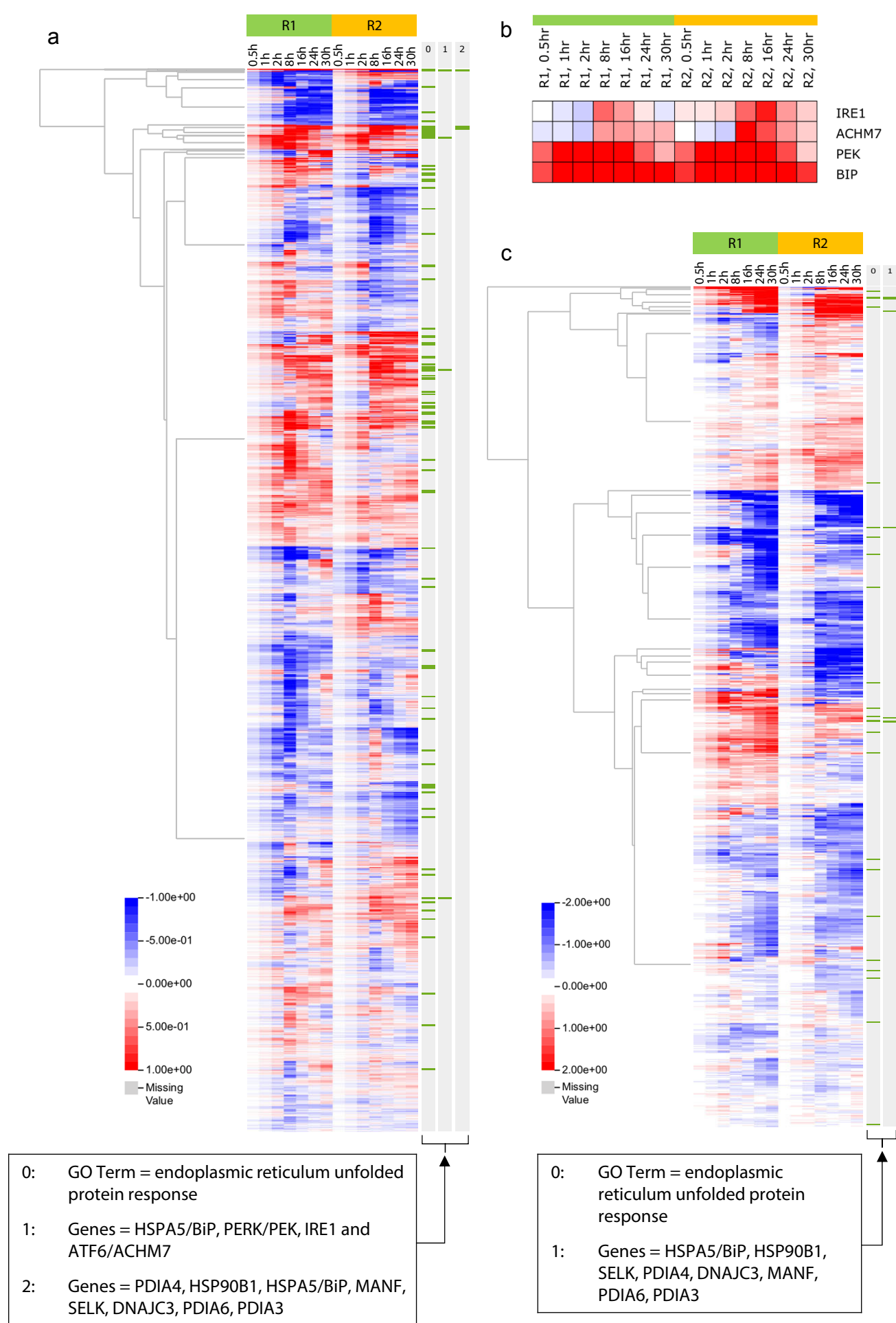
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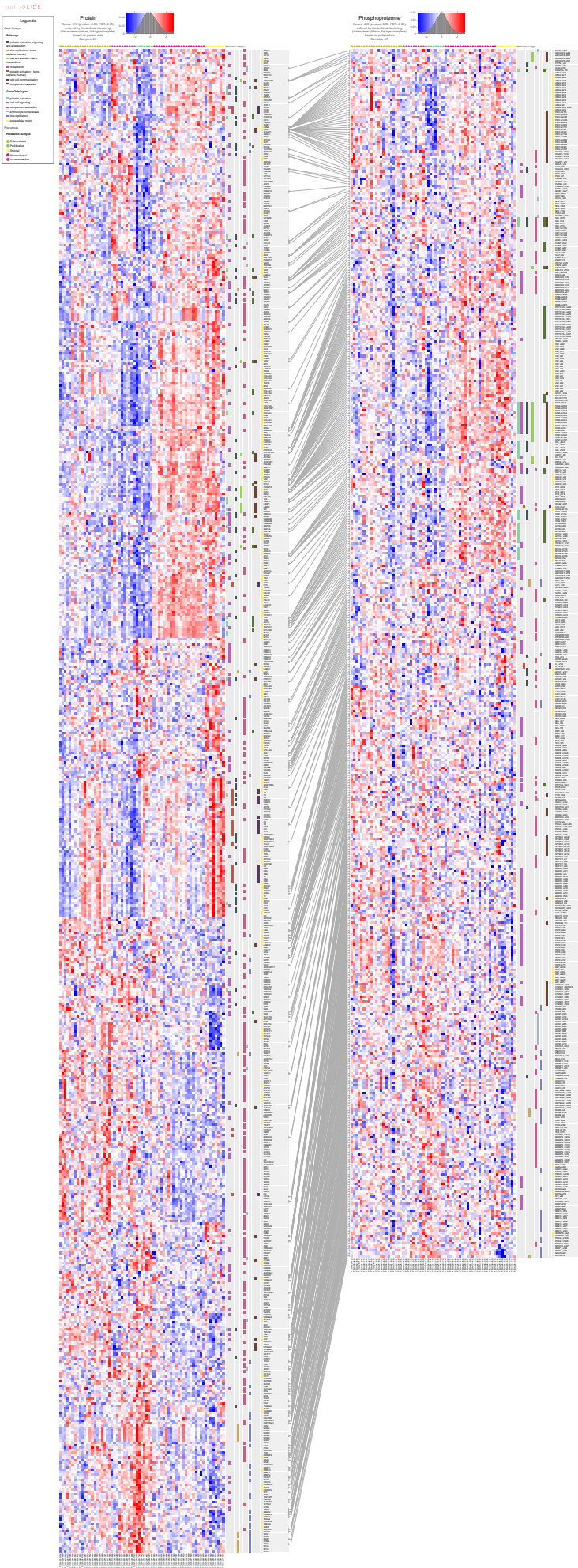
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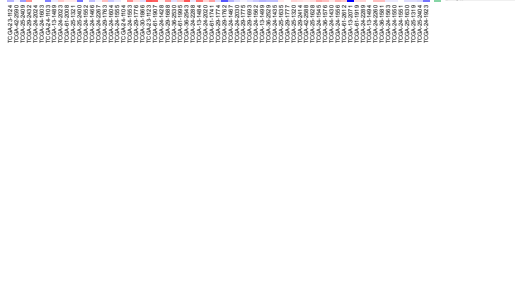
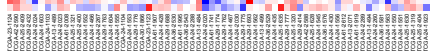
Supplementary Fig. S1: Visualization interface of multiSLIDE. a. The main visualization home for multiSLIDE. On the top-left corner is the “Add Genes” button to search and add pathways and GO terms for visualization. Clicking this button opens the search panel shown in b. The three tabs in this panel, Search, Enrichment Analysis, and Upload, are respectively, for keyword-based pathway search, pathway enrichment analysis, and for uploading externally curated pathways. The menu panel at the top has options to save visualizations and analysis workspace, as well as for creating custom networks to visualize inter-omics relationships, as shown in d and e, respectively. The global settings panel, just below the menu panel, provides all the data exploration functionalities, such as joint (synchronous) or independent (asynchronous) clustering, data sorting and filtering functionalities, as shown in f. Each heatmap can be individually customized by clicking the ‘Customize Heatmap’ option beside the heatmaps, which opens the panel shown in c.



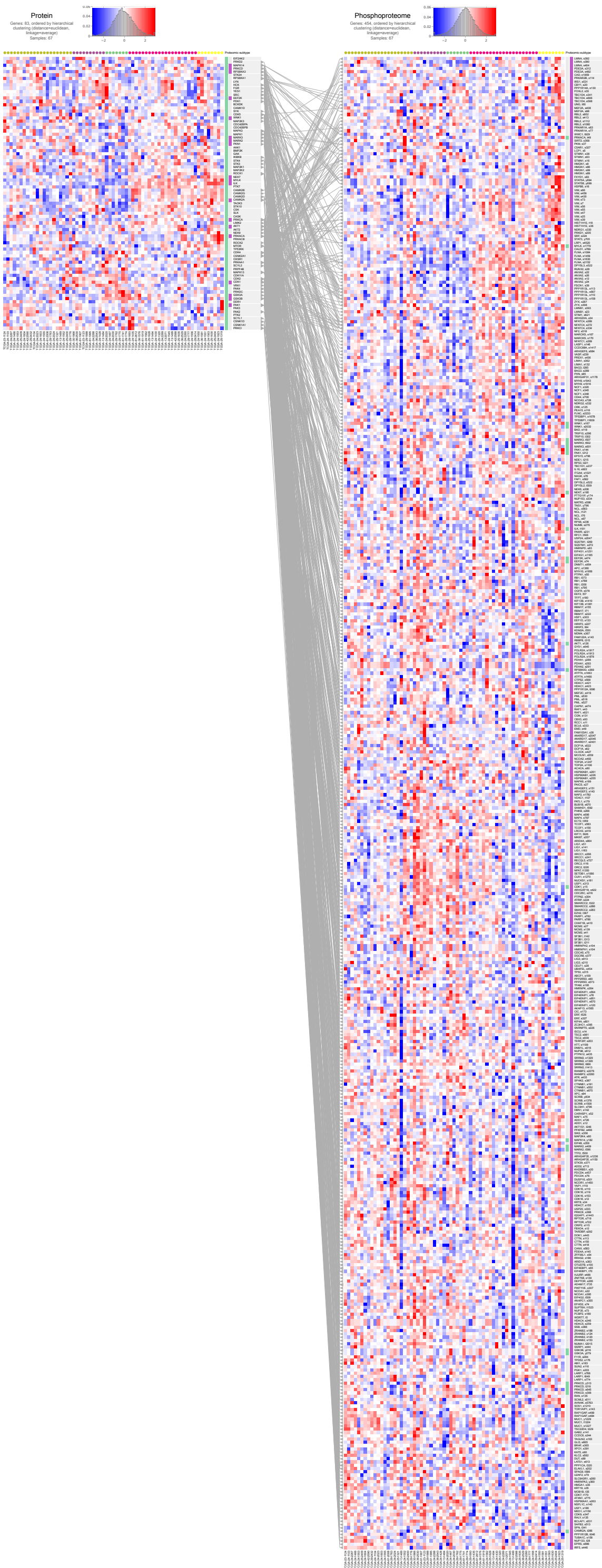
Supplementary Fig. S2: Visualization of individual omics data sets using SLIDE¹. **a.** The whole transcriptome data across the two replicates (N=16704), visualized using SLIDE's global heatmap view. The first search result highlights genes belonging to the GO term: endoplasmic reticulum (ER) unfolded protein response. The second search result highlights the four major sensor proteins in endoplasmic reticulum stress: HSPA5/BiP, PERK/PEK, IRE1, and ATF6/ACHM7. The third search result shows some known ER stress related and resident proteins. **b.** Visualizes the transcriptome level expression for the four sensor proteins separately. **c.** Visualizes the protein expression data after statistical filtering across the two replicates (N=1237) in SLIDE. The first search result highlights proteins belonging to the GO term: endoplasmic reticulum unfolded protein response. Both -omics data were baseline transformed by subtracting their respective 0 h time points (in log scale, base 2). The heatmaps show three phases of the stress response early (< 2 h), intermediate (2 - 8 h), and late (> 8 h). The whole mRNA regulation suggests a spike-like pattern in the transition from the early phase to the intermediate phase of the stress response, peaking in the intermediate phase before returning to original levels or stabilizing to a new equilibrium state different from 0h in the late phase. The protein level visualization shows a delayed response that persisted till the late stage (30 h).



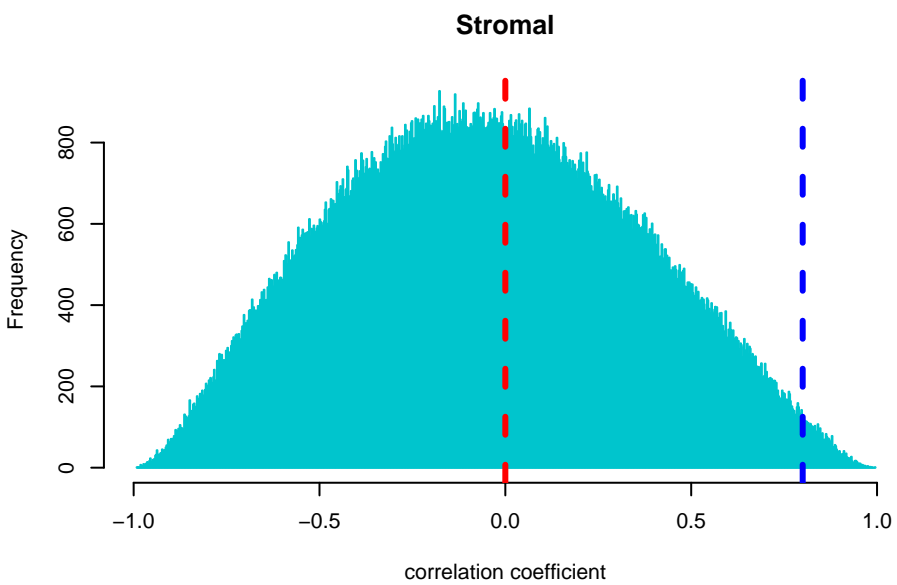
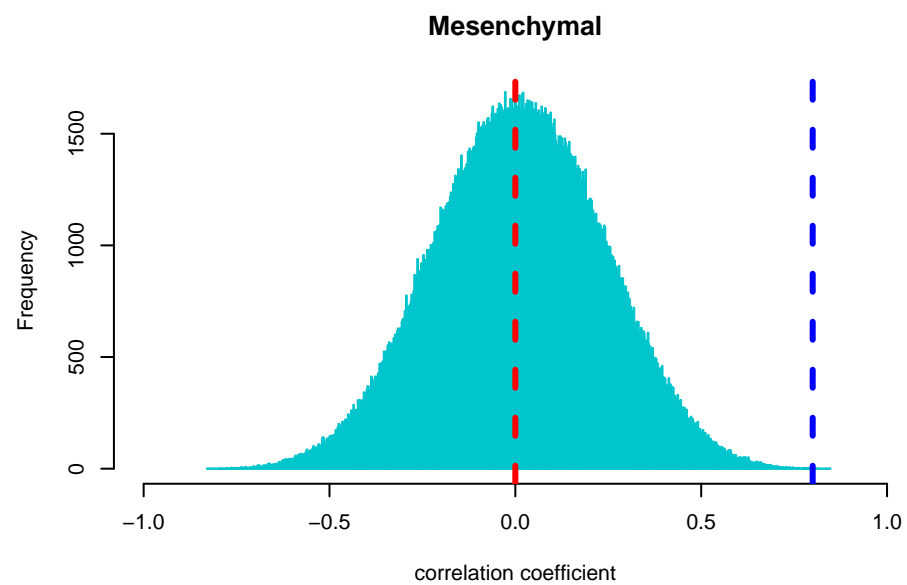
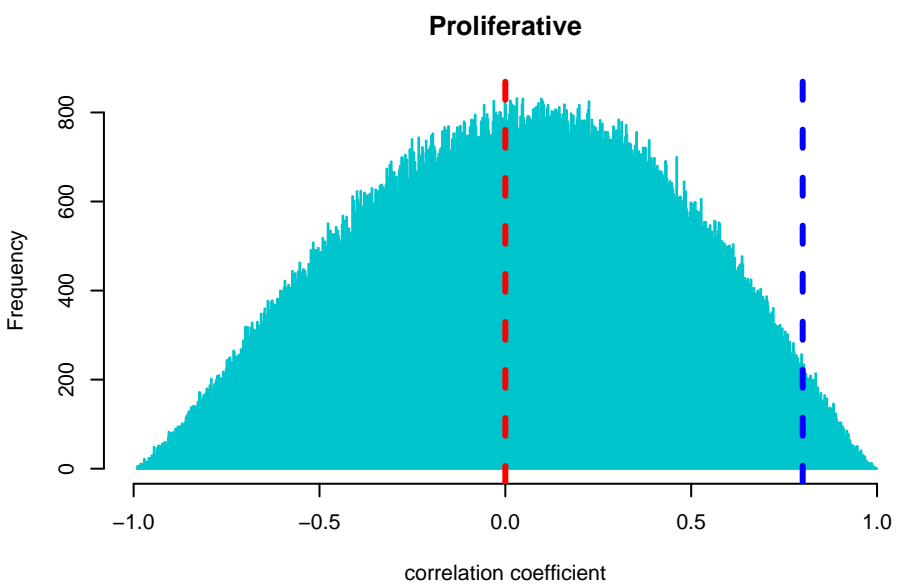
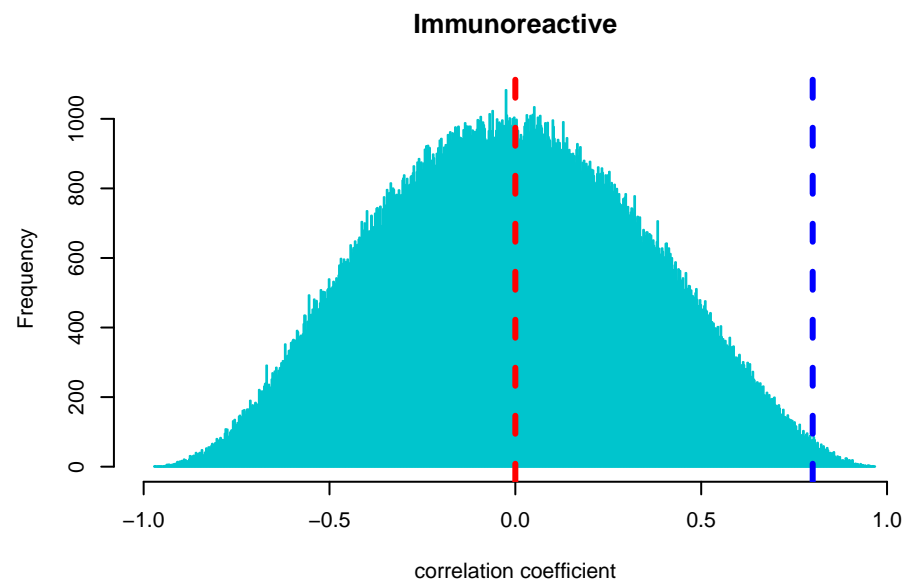
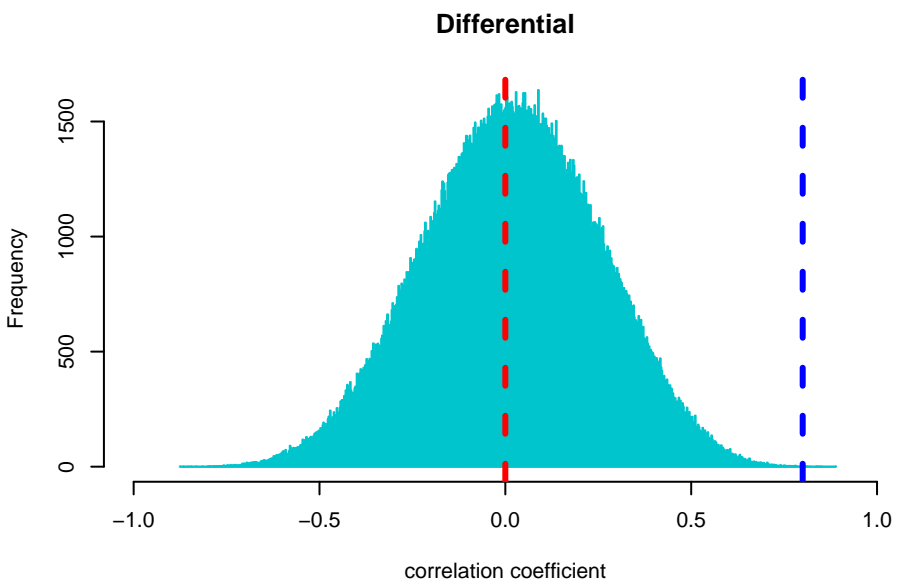
Supplementary Fig. S3: Pathway-based visualization of proteome and phosphoproteome data from the CPTAC Ovarian Cancer Cohort. Here, the proteome-based molecular subtypes: Differentiated, Immunoreactive, Proliferative, Mesenchymal, and Stromal, are used to group samples in both the proteome and phosphoproteome data. We performed keyword-based pathway or GO term searches following the pathway enrichment analysis by Zhang et al.² The legend panel shows the list of pathways and GO terms included in the visualization. Differential expression analysis using multiSLIDE's filtering feature identified 610 proteins and 490 phosphosites, using one-way ANOVA (p-value ≤ 0.05) for molecular subtypes and 5% FDR cutoff, as indicated beside the histograms. The heatmaps are synchronized, and hierarchical clustering of the protein data is used to determine the order. The visualization shows that there are similar expression patterns across the two omics levels, with phosphosites predominantly upregulated in the Mesenchymal and Stromal subtypes.



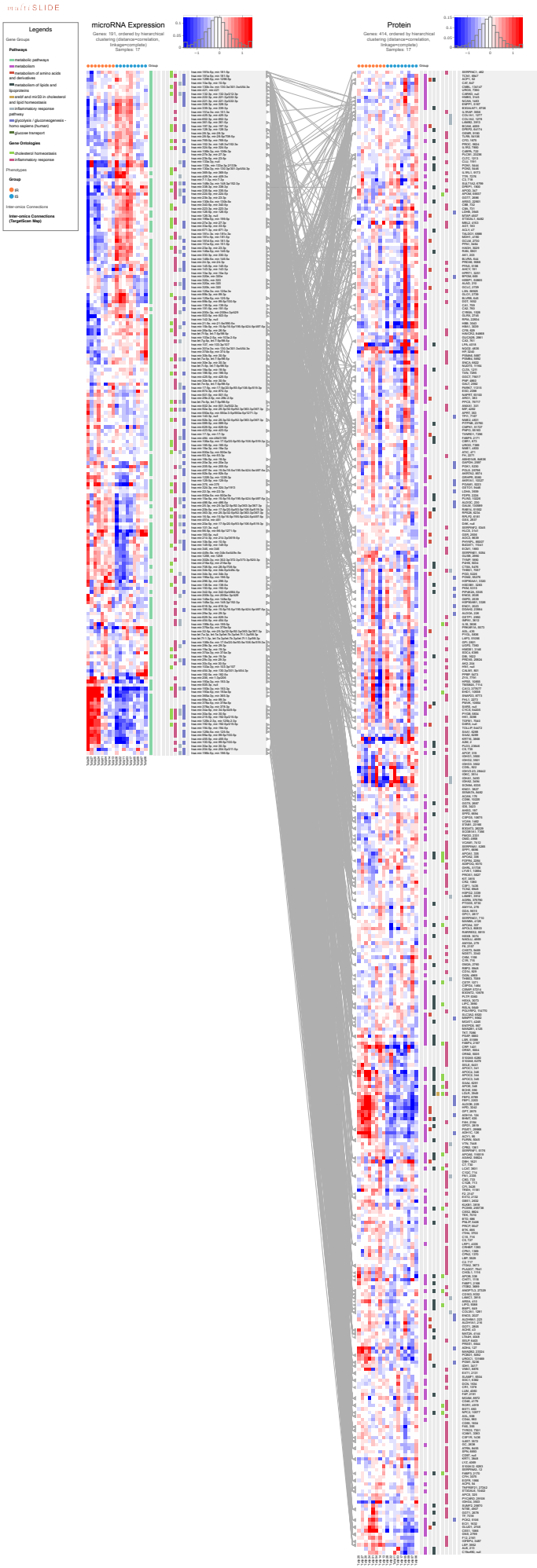
Supplementary Fig. S4: Visualization of the GO term ‘Extracellular Matrix’ in the CPTAC Ovarian Cancer Cohort. Genes belonging to the GO term “Extracellular Matrix” are visualized here. Differential expression analysis using multiSLIDE's filtering feature identified 155 proteins and 116 phosphosites. The filtering used one-way ANOVA (p-value ≤ 0.05) based on the molecular subtypes and 10% FDR cutoff. In contrast to **Supplementary Fig. S3**, here, proteins and phosphosites are clustered independently. The clusters in the protein data are highlighted using green bars. The visualization shows that proteins such as Plectin (PLEC), Lamin A/C (LMNA), Filamin A (FLNA), and Vimentin (VIM) have the highest number of sites that are differentially phosphorylated, with consistent patterns with the differences in the protein abundance.



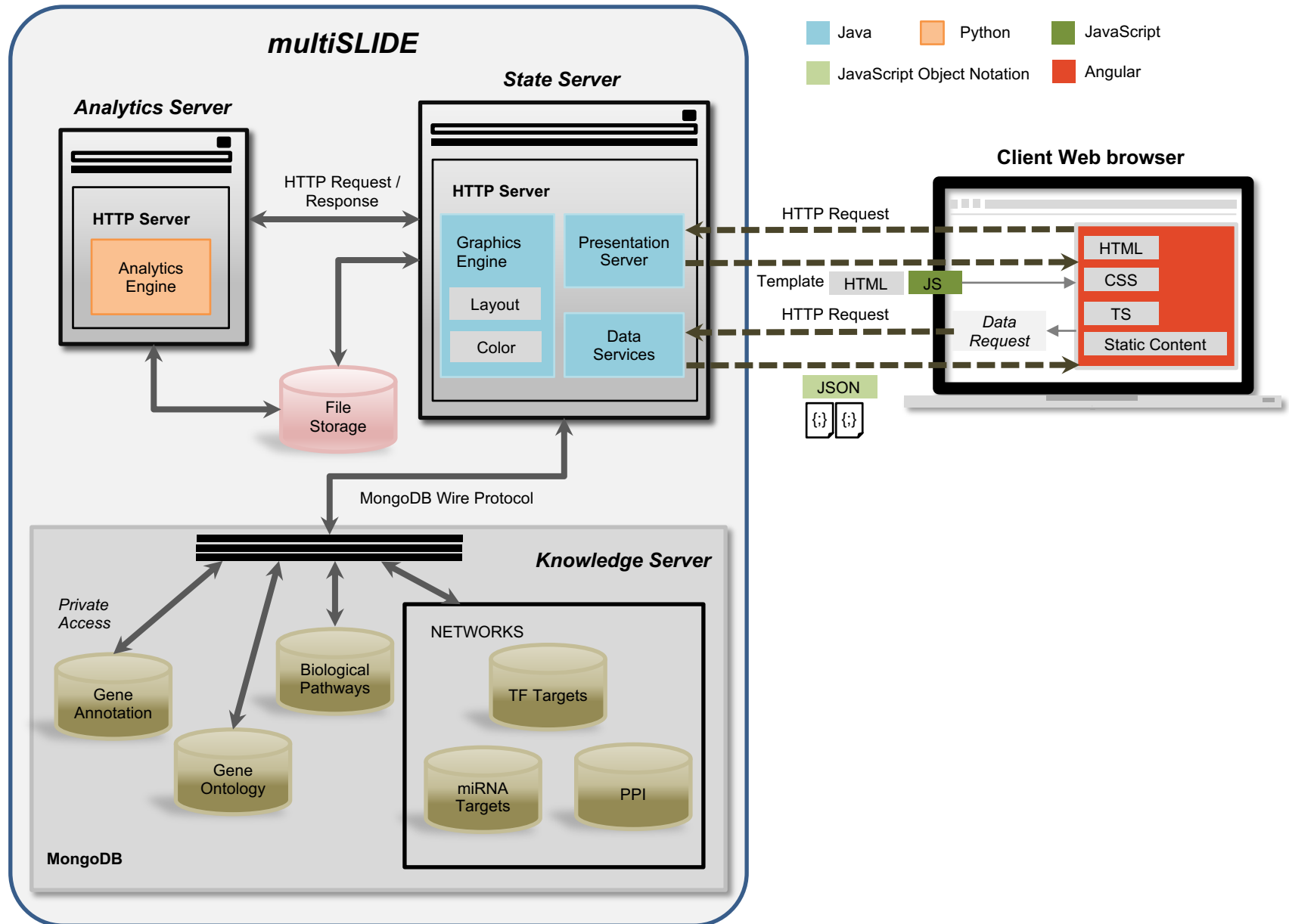
Supplementary Fig. S5: Visualization of kinase-substrate relationship in the CPTAC Ovarian Cancer Cohort. The figure visualizes kinases in the proteome data and their corresponding substrates in the phosphoproteome data. We curated kinase-substrate interactions from PhosphoSitePlus,³ PhosphoNetworks,⁴ and a predictive network inference approach⁵ to build a kinase-substrate map. All kinase-substrate pairs present in both the kinase-substrate map and in the datasets are visualized here. **Fig. 3** shows a subset of these kinases-substrate pairs, where the proliferative subtype is upregulated.



Supplementary Fig. S6: Distribution of correlation coefficients between kinase-site pairs in the CPTAC Ovarian Cancer Cohort. The five panels show the distribution of the Pearson correlation coefficients between the subtype-specific expressions of kinase-site pairs for the five molecular subtypes in the CPTAC Ovarian Cancer Cohort. The correlation coefficients are calculated between the 83 kinases visualized in **Supplementary Fig. S5** and all phosphosites. The blue vertical line indicates a correlation value of 0.8. For the Immunoreactive, Proliferative, and Stromal subtypes, a large number of positively correlated kinase-site pairs are present. We also see in **Supplementary Fig. S5** that several kinases are upregulated in the Proliferative subtype. Therefore, in **Fig. 3**, we focus on kinase-site pairs that are highly correlated and show upregulation in the Proliferative subtype.



Supplementary Fig. S7: Visualization of human plasma proteome and microRNA-ome associated with insulin resistance. The figure shows pathways selected by searching for: metabolic pathways, metabolism, inflammatory response, glucose transport, and lipid homeostasis. Here, proteins and miRNA are independently clustered using correlation distance (1 minus Pearson correlation) and complete linkage function. The relationships between miRNA family names and their target proteins are extracted from TargetScanMap and uploaded into multiSLIDE using the network upload feature. A subset of these proteins and miRNA, identified through differential expression analysis, are visualized in **Fig. 4**.



Supplementary Fig. S8: Software architecture of multiSLIDE. The figure presents a schematic of multiSLIDE's software architecture. multiSLIDE is built on a distributed architecture with the server comprising of three components: a state server, an analytics server, and a knowledge server. The client can be any modern web browser. The state and analytics servers are HTTP servers, with the state server maintaining client state information and holding uploaded data, whereas the analytics server is the main computation engine. The knowledge server implemented using MongoDB, manages the physical storage of curated gene annotation, regulatory networks, biological pathways, and Gene Ontologies (GO). The analytics and knowledge servers are stateless. The client interacts only with the state server.

List of abbreviations: HTTP – Hypertext Transfer Protocol, HTML – Hypertext Markup Language, TF – Transcription Factor, PPI – Protein-Protein Interaction, JS – JavaScript, CSS – Cascading Style Sheets, JSON – JavaScript Object Notation, TS – TypeScript.

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